Datasheet

Revision date: 08 Oct



Nitric Oxide Synthase, Inducible (NOS2) Antibody Pair

Catalogue No.:BTA100025

Nitric Oxide Synthase 2, Inducible (NOS2) Antibody Pair for use in Sandwich ELISA assay development. This antibody pair contains:

Component 5 × 96 tests 10 × 96 tests

Capture Antibody 200 μg 400 μg Biotin-Conjugated Detection Antibody50 μg 100 μg Standard 2 μg 10 μg

Please note that quantities and concentrations may change between different batches.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method)

Target: Nitric Oxide Synthase, Inducible (NOS2)

Reactivity: Mouse

Tested Applications: ELISA

Recommended dilutions: Dilute the Capture Antibody 125-fold with Coating Buffer.

Dilute the Biotin-Conjugated Detection Antibody 200-fold with Detection Antibody Diluent.

Optimal dilutions/concentrations should be determined by the end user.

Form: Liquid (Capture Antibody and Detection Antibody)

Reconstitution: Reconstitute the standard with Standard Diluent. The volume, and therefore standard

concentration, should be determined by the end user.

Storage: Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles.

Buffer: The Capture and Detection Antibody both contain 0.1% sodium azide.

Standard Form: Lyophilized

Assay Type: Sandwich

Capture Antibody Conjugation: Unconjugated

Detection Antibody Conjugation:Biotin

Concentration: Capture Antibody: 0.5 mg/ml

Biotin-Conjugated Detection Antibody: 0.2 mg/ml

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Note:

This product is for research use only.

Directions for use:

Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

- Dilute the Capture Antibody to working concentration using Coating Buffer.
 Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well).
 Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
 - 2. Aspirate the wells and wash with Wash Buffer (350 μl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
- 2. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
- 3. Repeat the aspiration/wash process in Step 2.
- 4. Add 100 μl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
- 5. Repeat the aspiration/wash process in Step 2.
- Add appropriately diluted Biotin-Conjugated Detection Antibody (100 μl per well).
 Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
- 7. Repeat the aspiration/wash process in Step 2.
- 8. Add appropriately diluted Streptavidin HRP (100 μl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
- 9. Repeat the aspiration/wash process in Step 2.
- Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
- 11. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
- 12. Measure the absorbance immediately using a microplate reader set at 450 nm.