Immunoglobulin G1 (IgG1) Antibody Pair

Catalogue No.:BTA10005

Immunoglobulin G1 (IgG1) Antibody Pair for use in Sandwich ELISA assay development. This antibody pair contains:

Component	5 × 96 tests	10 × 96 tests
Capture Antibody	200 µg	400 µg
Biotin-Conjugated Detection Antibody	50 µg	100 µg
Standard	2 µg	10 µg

Please note that quantities and concentrations may change between different batches.

It is recommended to use this antibody pair with abx098958 Antibody Pair Support Kit (Sandwich Method).

Target: Immunoglobulin G1 (IgG1)

Reactivity: Human

Tested Applications: ELISA

Recommended dilutions: Dilute the Capture Antibody 125-fold with Coating Buffer.
Dilute the Biotin-Conjugated Detection Antibody 200-fold with Detection Antibody Diluent.
Optimal dilutions/concentrations should be determined by the end user.

Form: Liquid (Capture Antibody and Detection Antibody)

Reconstitution: Reconstitute the standard with Standard Diluent. The volume, and therefore standard concentration, should be determined by the end user.

Storage: Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles.

Buffer: The Capture and Detection Antibody both contain 0.1% sodium azide.

Standard Form: Lyophilized

Assay Type: Sandwich

Capture Antibody Conjugation: Unconjugated

Detection Antibody Conjugation: Biotin

Concentration: Capture Antibody: 0.5 mg/ml
Biotin-Conjugated Detection Antibody: 0.2 mg/ml

Note: This product is for research use only.

Directions for use: Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

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1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 4 °C overnight or at 37 °C for 2 hours
   2. Aspirate the wells and wash with Wash Buffer (350 µl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper.
2. Block the plate with Blocking Buffer (200 µl per well) at 37 °C for 1.5 hours.
3. Repeat the aspiration/wash process in Step 2.
4. Add 100 µl of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1 hour.
5. Repeat the aspiration/wash process in Step 2.
6. Add appropriately diluted Biotin-Conjugated Detection Antibody (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
7. Repeat the aspiration/wash process in Step 2.
8. Add appropriately diluted Streptavidin HRP (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
9. Repeat the aspiration/wash process in Step 2.
10. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 10-20 min. Keep the plate in the dark and avoid exposure to light.
11. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
12. Measure the absorbance immediately using a microplate reader set at 450 nm.